## New Methoxytriterpene Dione from the Cuticle of Picea jezoensis var. jezoensis

Reiko Tanaka, Kazuhiro Tsujimoto, Yasuko In, and Shunyo Matsunaga\*

Osaka University of Pharmaceutical Sciences, 4-20-1 Nasahara, Takatsuki, Osaka 569-11, Japan

Osamu Muraoka and Toshie Minematsu

Faculty of Pharmaceutical Sciences, Kinki University, 3-4-1 Kowakae, Higashiosaka, Osaka 577, Japan

Received August 30, 1996<sup>®</sup>

A novel pentacyclic triterpene dione was isolated from the cuticle of *Picea jezoensis* var. *jezoensis* together with the known serrat-14-ene-3,21-dione (**1**), and the structure of this compound was determined as  $21\alpha$ -methoxyserrat-13-ene-3,15-dione (**2**). Detailed NOESY experiments revealed that **2** has a chair form of ring A and a chairlike conformation of ring C, respectively, in CDCl<sub>3</sub> solution. Interestingly, single-crystal X-ray analysis indicates that in the solid state **2** has a deformed boat form of ring A, in which the 3-oxo and the 25-methyl groups are arranged in flag-pole positions, and a chairlike form of ring C.

Previously, we reported that the CH<sub>2</sub>Cl<sub>2</sub> extract of the cuticle of *Picea jezoensis* (Sieb. et Zucc.) Carr. var. *jezoensis* Mayr. (Pinaceae) contained  $21\alpha$ -hydroxy- $3\beta$ -methoxyserrat-14-en-21-one, and  $14\beta$ ,  $15\beta$ -epoxy- $3\beta$ -methoxyserrat-14-en-21 $\beta$ -ol, together with eight known triterpenoids.<sup>1,2</sup> Further work on other constituents of the cuticle has now led to the isolation of a new unsaturated triterpene dione (**2**), along with the known compound, serrat-14-en-3,21-dione (**1**).<sup>3-5</sup> Herein, we describe the structure of **2**.



The known compound was confirmed as serrat-14ene-3,21-dione (1), which had been synthesized from serratenediol<sup>3</sup> and later isolated from the bark of *Pinus*  *luchuensis* Mayer.<sup>4</sup> Physical and spectral data of **1** were in good agreement with those already reported, while the <sup>13</sup>C-NMR spectrum is reported here for the first time. Detailed assignments for the <sup>13</sup>C-NMR signals are given in Table 1, together with the <sup>1</sup>H-NMR signals.

Compound 2 was assigned the molecular formula of C<sub>31</sub>H<sub>48</sub>O<sub>3</sub> by HREIMS. The UV and IR spectra exhibited absorption bands characteristic for an  $\alpha$ , $\beta$ -unsaturated ketone and a saturated six-membered ring ketone  $(\lambda_{\text{max}} 230 \text{ and } 282 \text{ nm}; \nu_{\text{max}} 1654 \text{ and } 1703 \text{ cm}^{-1})$ . In the <sup>1</sup>H- and <sup>13</sup>C-NMR spectra (Table 1), 2 exhibited signals for seven quaternary methyl groups, two methylene groups [ $\delta_{\rm H}$  2.36–2.48 (4H)] vicinal to two different carbonyl carbons [ $\delta_{\rm C}$  199.0 (s) and 217.9 (s)], and an equatorially oriented secondary methoxy group [ $\delta_{\rm H}$  2.76 (1H, dd, J = 11.8, 4.0 Hz, H-21 $\beta$ ) and 3.39 (3H, s),  $\delta_{\rm C}$ 57.6 (q) and 87.4 (d)]. In spite of the absence of any olefin proton signals, 2 showed <sup>13</sup>C-NMR signals for two sp<sup>2</sup> quaternary carbons at  $\delta_{\rm C}$  133.0 and 169.8. Thus, it became obvious that 2 must have a tetrasubstituted double bond conjugated to one of two keto groups in the molecule. In addition, the <sup>1</sup>H-NMR signals appeared as an AB system at rather low magnetic field at  $\delta_{\rm H}$  1.72 and 2.81 (each 1H, d, J = 14.5 Hz). These signals were attributable to the isolated 27-methylene group in the serratene skeleton.<sup>6</sup> Analysis of all these data suggested that the structure of 2 was an unknown methoxyserratene dione including either the chromophore of a  $\Delta^{13}$ -en-12-one or a  $\Delta^{13}$ -en-15-one. The <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C COSY and COLOC experiments performed on **2** supported the latter structure. The COLOC data provided cross-correlations as shown in Table 1. These correlations indicated 2 to have a gross carbon framework that was presumed to be that of 21-methoxyserrat-13-ene with attached keto groups at C-3 and C-15.

Compound **2** showed nine significant fragment peaks corresponding to ions  $\mathbf{a}-\mathbf{i}$  between 315 and 121 atomic mass units in its EIMS, along with the parent ion peak (see Experimental Section). Of these, ions  $\mathbf{a}-\mathbf{d}$  and  $\mathbf{g}$  are characteristic for the fragmentation of a serrat-13-ene skeleton bearing a keto group at C-15. Together with the presence of an equatorial secondary methoxy proton signal, these data indicated compound **2** to be 21 $\alpha$ -methoxyserrat-13-ene-3,15-dione.

<sup>\*</sup> To whom correspondence should be addressed. Phone: 81-726-90-1084. FAX: 81-726-90-1005.

<sup>&</sup>lt;sup>®</sup> Abstract published in Advance ACS Abstracts, March 1, 1997.

T 11 4			<b>C</b> 1	4 10	2
l'able I.	IN M R.	Data for	Compounds	$\mathbf{I}$ and $\mathbf{Z}$	2

Notes

	$\delta_{ m H}$		$\delta_{\mathrm{C}}$			
position	1	2	1	2	COLOC data for 2 (C to H)	
1	1.49 m	1.54α m	39.5 t	39.2 t	H-5α, H-9α, Me-25	
	2.03 m	1.95β m				
2	2.46 m	2.47a m	34.1 t	33.9 t		
	2.48 m	2.48β m				
3		1	218.2 s	217.9 s	H-5α, Me-23, Me-24	
4			47.3 s	47.3 s	Me-23, Me-24	
5	1.42 dd (13.8, 2.5)	1.53α m	55.2 d	54.9 d	Me-23, Me-24, Me-25	
6	1.46 m	1.48α m	20.1 t	20.1 t		
	1.57 m	1.54β m				
7	1.26 m	1.36a td (12.5, 3.9)	44.2 t	43.0 t	H-5α, H-9α, Me-26	
	1.42 m	1.63β m				
8		1	37.0 s	34.9 s	Me-26	
9	0.93 br s	1.14α br s	62.1 d	63.8 d	H-1, H-5α, H-7, H-27, Me-25, Me-26	
10			37.8 s	37.9 s	H-11, Me-25	
11	1.12 m	1.81β m	25.5 t	20.6 t		
	1.76 m	2.07a m				
12	1.96 m	2.58α dd (14.0, 8.2)	27.7 t	29.7 t	Η-9α	
	1.22 m	<b>2.06</b> β m				
13	1.81 dd (15.1, 1.3)	1	56.5 d	169.8 s	H-11, H-17β, H-27, Me-28	
14			137.9 s	133.0 s	H-12	
15	5.40 dif. t (2.9)		122.3 d	199.0 s	$H-17\beta$	
16	2.02 m	2.36α dd (12.7, 14.2)	24.5 t	34.6 t		
	2.06 m	$2.49\beta$ dd (14.2, 3.5)				
17	1.67 dd (12.5, 3.4)	$1.63\beta$ dd (12.7, 3.5)	51.2 d	50.1 d	Me-28, Me-29, Me-30	
18			36.2 s	40.4 s	Me-28	
19	1.43 m	1.44β m	38.4 t	34.1 t	H-17β, Me-28	
	2.17 ddd (12.9, 5.3, 3.8)	2.03a dt (14.8, 4.9)				
20	2.46 dt (14.2, 3.8)	1.59β m	34.8 t	22.3 t		
	2.76 td (14.2, 5.3)	1.99a m				
21		2.76β dd (11.8, 4.0)	216.9 s	87.4 d	Me-29, Me-30, OMe	
22			47.7 s	38.5 s	Me-29, Me-30	
23	1.08 s	1.09 s	26.9 q	26.9 q	H-5a, Me-24	
24	1.04 s	1.03 s	21.0 q	20.3 q	H-5a, Me-23	
25	0.89 s	0.83 s	15.8 q	16.2 q	Η-1, Η-5α	
26	0.87 s	0.69 s	19.3 q	18.7 q	Η-7, Η-9α, Η-27	
27	1.84 d (14.6)	1.72α d (14.5)	55.8 t	40.6 t	H-7, H-9a, Me-26	
	2.26 d (14.6)	2.81β d (14.5)				
28	0.93 s	1.04 s	13.0 q	17.1 q	H-17β, H-19	
29	1.05 s	0.86 s	24.5 q	27.6 q	H-17 $\beta$ , H-21 $\beta$ , Me-30	
30	1.09 s	1.00 s	24.5 g	27.6 a	H-17 $\beta$ , H-21 $\beta$ , Me-29	
OMe		3.39 s	I	57.6 q	H 21 $\beta$	

<sup>a</sup> The <sup>1</sup>H- and <sup>13</sup>C-NMR correlations were based on HETCOR spectra.



Figure 1. ORTEP diagram of 2.

Previous workers have examined the CD spectrum and single-crystal X-ray analytical data for serrat-14ene-3,21-dione (1) and reported this molecule to have a deformed boat form of ring A and a chairlike conformation of ring C.<sup>5</sup> Therefore, compound **2** provided a stereochemical uncertainty that needed to be resolved, inasmuch as it contains a  $\Delta^{13}$ -en-15-one grouping and the same 4,4-dimethyl-3-one system as **1** in the molecule. The single-crystal X-ray analysis furnished an ORTEP diagram for the structure of **2** (Figure 1), in which rings A and C adopt a deformed boat conformation so as to keep the 3-oxo and the 25-methyl groups on the flag-pole positions of the ring and a chairlike



form, respectively, when in the solid state. Table 2 shows the list of nonhydrogen atom fractional coordinates for 2.

On the other hand, the NOESY spectrum of **2** (Figure 2) exhibited correlations for the signals of Me-26 (with H-6 $\beta$ , H-7 $\beta$ , H-11 $\beta$ , and H-27 $\beta$ ), H-7 $\alpha$  (with H-27 $\alpha$ ), H-7 $\beta$  (with H-27 $\beta$ ), Me-28 (with H-12 $\alpha$ , H-16 $\alpha$ , and H-20 $\alpha$ ), H-21 $\beta$  (with H-19 $\beta$  and Me-30), Me-29 (with H-16 $\alpha$  and Me-28), Me-30 (with H-17 $\beta$  and H-21 $\beta$ ), H-9 $\alpha$  (with H-12 $\alpha$ ), and H-17 $\beta$  (with H-21 $\beta$  and Me-30). These data indicated that ring C of **2** is *trans*-fused with ring B to adopt a chairlike conformation. More significantly, cross interactions were observed for the signals of Me-

**Table 2.** Non-Hydrogen Atom Fractional Coordinates for Compound 2 (Esd's in parentheses)

-				
atom	X	У	Z	U(eq)
O(1)	0.1346(7)	0.6121(2)	0.3404(8)	0.105(4)
O(2)	0.0303(5)	0.8425(2)	1.2493(6)	0.088(3)
O(3)	-0.0093(4)	1.0014(1)	0.5813(7)	0.077(3)
C(1)	0.2746(5)	0.7031(2)	0.5077(8)	0.053(3)
C(2)	0.2870(7)	0.6582(2)	0.440(1)	0.071(4)
C(3)	0.1795(7)	0.6314(2)	0.463(1)	0.070(4)
C(4)	0.1275(6)	0.6293(2)	0.6507(9)	0.059(3)
C(5)	0.1573(5)	0.6704(2)	0.7532(7)	0.050(3)
C(6)	0.0732(6)	0.6766(2)	0.9103(8)	0.061(3)
C(7)	0.1116(6)	0.7140(2)	1.0193(7)	0.058(3)
C(8)	0.1134(4)	0.7554(2)	0.9160(6)	0.046(3)
C(9)	0.1950(4)	0.7487(2)	0.7519(6)	0.044(3)
C(10)	0.1657(4)	0.7097(2)	0.6342(6)	0.043(2)
C(11)	0.2147(4)	0.7885(2)	0.6422(7)	0.046(3)
C(12)	0.2819(4)	0.8237(2)	0.7386(8)	0.053(3)
C(13)	0.2012(4)	0.8512(2)	0.8526(8)	0.052(3)
C(14)	0.1522(5)	0.8331(2)	0.9969(7)	0.052(3)
C(15)	0.0773(5)	0.8585(2)	1.1191(8)	0.062(3)
C(16)	0.0632(7)	0.9033(2)	1.0785(9)	0.068(4)
C(17)	0.0637(5)	0.9116(2)	0.8821(8)	0.053(3)
C(18)	0.1846(5)	0.8963(2)	0.8017(7)	0.051(3)
C(19)	0.1725(6)	0.9018(2)	0.5978(8)	0.059(3)
C(20)	0.1390(5)	0.9459(2)	0.5492(8)	0.064(3)
C(21)	0.0203(6)	0.9596(2)	0.6293(9)	0.058(3)
C(22)	0.0181(6)	0.9563(2)	0.8300(8)	0.059(3)
C(23)	-0.0066(7)	0.6172(2)	0.644(1)	0.076(4)
C(24)	0.1967(7)	0.5926(2)	0.740(1)	0.087(5)
C(25)	0.0517(5)	0.7161(2)	0.5235(7)	0.049(3)
C(26)	-0.0152(5)	0.7698(2)	0.8762(7)	0.055(3)
C(27)	0.1710(5)	0.7888(2)	1.0422(7)	0.054(3)
C(28)	0.3000(5)	0.9190(2)	0.865(1)	0.071(4)
C(29)	0.0918(7)	0.9919(2)	0.912(1)	0.083(5)
C(30)	-0.1146(7)	0.9620(2)	0.892(1)	0.081(5)
C(31)	-0.0626(8)	1.0059(3)	0.414(1)	0.089(5)



Figure 2. NOESY correlations of compound 2.

23 (with H-5 $\alpha$ ), Me-24 (with H-2 $\beta$ ), Me-25 (with H-1 $\beta$ , H-2 $\beta$ , and H-11 $\beta$ ), and H-1 $\alpha$  (with H-5 $\alpha$ ). Contrary to our expectation, the above results indicated that ring A of compound 2 was trans-fused with ring B and adopts a chair form in CDCl<sub>3</sub>, as Me-25 showed extremely clear NOESY enhancements with Me-24 and H-2 $\beta$ . In four methylene proton signals appearing between  $\delta$  2.33 and 2.51, those resonating at  $\delta$  2.36 (1H, dd, J = 12.7, 14.2Hz) and 2.49 (1H, dd, J = 14.2, 3.5 Hz) were assigned to H-16 $\alpha$  and H-16 $\beta$ , respectively, on the basis of the <sup>1</sup>H-<sup>1</sup>H COSY and <sup>1</sup>H-<sup>13</sup>C COSY spectra. Similarly, the other two methylene proton resonances were observed at  $\delta$  2.47 and 2.48 (each 1H, m) and attributed to H-2 $\alpha$ and H-2 $\beta$ , respectively. In addition, methylene signals observed at  $\delta$  2.58 (1H, dd, J = 14.0 and 8.2 Hz) and 2.06 (1H, m) were assigned, in turn, to H-12 $\alpha$  and H-12 $\beta$ . A methine proton at  $\delta$  1.63 (1H, dd, J = 12.7, 3.5 Hz) was assigned to H-17 $\beta$ . Although an attempt to prove the exact conformation by analyzing the coupling constants of the H-2 $\alpha$  and H-2 $\beta$  signals was unsuccessful (because these signals overlapped each other along with those of H-16 $\beta$ ), the  $\delta$  values and signal patterns of H-2 $\alpha$ , H-2 $\beta$ , and H-17 $\beta$  for **2** were closely similar when compared with those of **1**. Concerning rings A and B, **1** showed the same NOESY correlations as those of **2**, as mentioned above. Hence, **1** must also have a chair form of ring A in CDCl<sub>3</sub>.

The above results demonstrate that in compounds **1** and **2** ring A changes its conformation according to whether it is in the solid state or in solution. Therefore, in the former state it adopts a defomed boat form, whereas it takes a chair form in the latter state.

## **Experimental Section**

General Experimental Procedures. Melting points were determined on a Ishii hot-stage apparatus and are uncorrected. Optical rotations were measured in CHCl<sub>3</sub> using a JASCO DIP-140 digital polarimeter. UV spectra were recorded in EtOH on a Hitachi model 150-20 spectrophotometer. IR spectra were run as KBr disks on a Perkin-Elmer 1720X FTIR spectrophotometer. <sup>1</sup>Hand <sup>13</sup>C-NMR spectra were obtained on a JEOL GX-500 spectrometer with standard sequences operating at 500 MHz and 125 MHz, respectively. CDCl<sub>3</sub> was used as solvent and TMS as internal standard. All <sup>13</sup>C-NMR assignments were based on DEPTGL, 2D <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H<sup>-13</sup>C COSY, COLOC, and NOESY experiments. EIMS and HREIMS were recorded on a Hitachi 4000H double-focusing mass spectrometer (70 eV). Column chromatography was carried out on Si gel (70-230 mesh, Merck). Fractions obtained from column chromatography were monitored by TLC (Si gel 60  $HF_{254}$ , 0.25-mm thick) with visualization under UV (254 and 365 nm). Preparative TLC was carried out on Merck Si gel  $PF_{254}$  plates (0.5-mm thick).

Extraction and Isolation. We have already reported that repeated column chromatography of residue **F**, one of six fractions separated from the  $CH_2Cl_2$  extract (365.1 g) of the cuticle of Picea jezoensis (Sieb. et Zucc.) Carr. var. jezoensis Mayr. (6.0 kg) by preliminary Si gel column chromatography, afforded  $14\beta$ ,  $15\beta$ -epoxy- $3\beta$ methoxyserratan-21-one (126 mg) and its  $21\beta$ -ol (107 mg) from the fractions eluted with CHCl<sub>3</sub> (fractions 39– 42, each fraction: 100 mL) and CHCl<sub>3</sub>-EtOAc (20:1, fractions 63-67), respectively.<sup>2</sup> At this stage, we had also collected 105 mg of an unexamined yellow gum showing two spots on TLC from the intermediary fractions of 43-61 eluted with CHCl<sub>3</sub> in the above separation. Further chromatography of the gummy product on 10% AgNO<sub>3</sub>-impregnated Si gel (10 g) using *n*-hexane $-C_6H_6$  (1:1) afforded a crystalline mass from fractions 15-21 (each fraction, 20 mL), which was purified by preparative TLC (solvent, CHCl<sub>3</sub>-MeOH, 100:1) to afford compound 2 (13 mg). Subsequent column chromatography with the same solvent furnished a crystalline solid from fractions 29-56, which was purified by preparative TLC to give compound 1: 40 mg, mp 204–206 °C (MeOH–CHCl<sub>3</sub>),  $[\alpha]^{23}_{D}$  –3.7° (c 0.43) [lit.<sup>3</sup> mp 208–210 °C,  $[\alpha]_D$  –3.7°]; IR (KBr)  $\nu_{max}$ 1714 (>C=O), 1637 and 807 (>C=CH-) cm<sup>-1</sup>; <sup>1</sup>H- and <sup>13</sup>C-NMR data, see Table 1; EIMS (70 eV) m/z [M]<sup>+</sup> 438, 423, 232, 218, 205, and 203. Physical and spectral data of 1 were in good agreement with those already published for serrat-14-ene-3,21-dione.<sup>3,4</sup>

**21**α-**Methoxyserrat-13-ene-3,15-dione (2)**: mp 317-319 °C (MeOH–CHCl<sub>3</sub>); [α]<sup>23</sup><sub>D</sub> +81.3° (*c* 0.28); HREIMS m/z 468.3602 [M]<sup>+</sup> (C<sub>31</sub>H<sub>48</sub>O<sub>3</sub> requires 468.3603); UV (EtOH)  $\lambda_{max}$  ( $\epsilon$ ) 230 (4300) and 282 (7000) nm (C=O and >C=C-C=O); IR (KBr)  $\nu_{max}$  2960, 2930, 2850, 1703 (C=O), 1654 (>C=C-C=O), 1465, 1455, 1419 (-CH<sub>2</sub>-CO), 1384 and 1363 (gem-dimethyl), 1179, 1149, 1106, 1033, 1010, 996, and 982 cm<sup>-1</sup>; <sup>1</sup>H- and <sup>13</sup>C-NMR data, see Table 1; EIMS (70 eV) m/z [M]<sup>+</sup> 468 (100), [M –  $Me^{+}_{+}$  453 (8), 439 (29),  $[M - MeOH]^{+}_{+}$  436 (4),  $[M - MeOH]^{+}_{+}$ - MeOH]<sup>+</sup> 421 (5), [ion **a**] 315.2320 [C<sub>21</sub>H<sub>31</sub>O<sub>2</sub>]<sup>+</sup> (99), [ion **b**] 299 (4), [ion **c**] 287.2004  $[C_{19}H_{27}O_2]^+$  (19), [ion **d**] 250.1927 [ $C_{16}H_{26}O_2$ ]<sup>+</sup> (100), [ion **e**] 219 (11), [ion **f**] 205 (15), [ion g] 203 (15), [ion h] 135 (20), [ion i] 121 (25).

X-ray Crystallography of Compound 2. A single crystal of 2 was obtained by recrystallization from a mixture of MeOH and CHCl<sub>3</sub>. Crystal data: C<sub>31</sub>H<sub>48</sub>O<sub>3</sub>, MW 468.72, orthorhombic, space group  $P2_12_12_1$ , a =11.067(2) Å, b = 31.969(2) Å, c = 7.586(1) Å, V =2683.8(5) Å<sup>3</sup>,  $D_x = 1.160$  g cm<sup>-3</sup>, Z = 4. A total of 2459 independent reflection intensities up to  $2\theta = 130^{\circ}$  were measured on a Rigaku four-circle diffractometer with graphite-monochromated CuKa radiation. A total of 1919 reflections with  $F_0 > 3\sigma F_0$  were used for the structure analysis by direct method. The non-hydrogen atoms were refined anisotropically by block-diagonal least-squares on a Micro-Vax computer at the Computer Center of Osaka University of Pharmaceutical Sciences. The geometrically ideal positions of H-atoms were calculated in the final refinement with isotropic thermal parameters; their electronic densities were ascertained on a difference Fourier map. The structure of compound **2** was finally refined to R = 0.059 ( $R_w = 0.085$ ). The

atomic scattering factors and anomalous scattering corrections were taken from International Tables for X-ray Crystallography.<sup>7</sup> All the crystallographic calculations were performed by the use of a CRYSTAN GM software package.<sup>8</sup> The final non-hydrogen atom fractional coordinates of 2 are listed in Table 2.9

Acknowledgments. This work was supported by the Section of Guidance and Spread, National Osaka Forestry Bureau, 1-8-75 Temmabashi, Kita-Ku, Osaka 530, Japan, for which the authors are deeply grateful. Our thanks are also due to Mrs. M. Fujitake of Osaka University of Pharmaceutical Sciences for MS measurements.

## **References and Notes**

- (1) Tanaka, R.; Senba, H.; Minematsu, T.; Muraoka, O.; Matsunaga, S. *Phytochemistry* **1995**, *38*, 1467–1471. (2) Tanaka, R.; Ohmori, K.; Minoura, K.; Matsunaga, S. *J. Nat.*
- Prod. 1996, 59, 237-241.
- Inubushi, Y.; Tsuda, Y.; Sano, T.; Konita, T.; Suzuki, S.; Ageta, H.; Otake, Y. *Chem. Pharm. Bull.* **1967**, *15*, 1153–1168.
- (4) Cheng, Y.-S.; Cheng, E. H.-T.; Fuang, G. J.-M. J. Chin. Chem. Soc. 1975, 22, 341–347.
- (5) Tsuda, Y.; Kashiwaba, N.; Hori, T. Chem. Pharm. Bull. 1983, 31, 1073-1076.
- (6) Seto, H.; Furihata, K.; Guangyi, X.; Xiong, C.; Deji, P. Agric.
- (7) Biol. Chem. 1988, 52, 1791–1801.
  (7) Ibers, J. A.; Hamilton, W. C. (Eds.). International Tables for X-ray Crystallography; The Keynoch Press: Birmingham, UK, 1974; Vol. IV.
- (8) CRYSTAN GM, A Computer Program for the Solution and Refinement of Crystal Structures from X-ray Diffraction Data, Version 6.1. MAČ Science Co. Ltd., 1984.
- (9) Atomic coordinates, thermal parameters, bond distance and angles observed and calculated structure factors of **2** have been deposited with the Cambridge Crystallographic Data Centre and can be obtained upon request from Dr. Olga Kennard, 12 Union Road, Cambridge CB2 1EZ, UK.

NP960604+